Supplementary Material for

Structure based optimization of designed Armadillo-repeat proteins.

Chaithanya Madhurantakam[#], Gautham Varadamsetty[#], Markus G. Grütter, Andreas Plückthun^{*} & Peer R. E. Mittl^{*}

Biochemisches Institut, Universität Zürich, Winterthurer Strasse 190, CH-8057, Zürich, Switzerland

PREM: Tel. +41-44-6356559, Fax. +41-44-6356834, e-mail. mittl@bioc.uzh.ch AP: Tel. +41-44-6355570, Fax. +41-44-6355712, e-mail. plueckthun@bioc.uzh.ch

[#] contributed equally to this work

| chains A and D | Hydrogen bonding | | chains B and C |
|----------------|------------------|------|----------------|
| residues) | Atom | Atom | |
| His4 | ND1 | OE1 | Glu114 |
| | ND1 | OE2 | Glu114 |
| His5* | NE2 | OE1 | Gln152 |
| | NE2 | NE2 | Gln152 |
| His6 | Ν | OE2 | Glu156 |
| | NE2 | OD1 | Asn121 |
| His7 | 0 | NE1 | Trp159 |
| His8 | NE2 | NE2 | Gln197 |
| His9 | Ν | OE2 | Glu198 |
| | NE2 | NE1 | Trp159 |
| | NE2 | ND2 | Asn163 |

Table S1. Hydrogen bonding interactions between the $\rm His_6\text{-}tag$ and the shallow concave groove on $Y_{II}M_4A_{II}.$

*His5 interacts only in chains A/B but not in chains C/D.

| Structure* | Repeats # | Curvature [†] | Twist [†] | Lateral bending ^{\dagger} |
|--|---|--|---|--|
| Y _{II} M ₄ A _{II} | M1/M2 M2/M3 M3/M4 M4/A5 average | $\begin{array}{c} 16.25 \pm 2.95 \\ 15.18 \pm 2.95 \\ 15.38 \pm 1.07 \\ 20.91 \pm 3.04 \\ 16.92 \end{array}$ | $\begin{array}{c} -22.32 \pm 1.65 \\ -23.63 \pm 0.74 \\ -23.16 \pm 0.50 \\ -28.18 \pm 3.43 \\ -24.32 \end{array}$ | $\begin{array}{c} -14.68 \pm 0.98 \\ -9.66 \pm 0.70 \\ -9.10 \pm 0.52 \\ -7.60 \pm 1.06 \\ -10.26 \end{array}$ |
| Y _{II} M ₃ A _{II} | M1/M2 M2/M3 M3/A4 average | $\begin{array}{c} 14.48 \pm 0.33 \\ 13.93 \pm 0.60 \\ 20.41 \pm 1.22 \\ 16.27 \end{array}$ | $\begin{array}{c} -22.39 \pm 0.79 \\ -22.98 \pm 0.71 \\ -26.09 \pm 2.75 \\ -24.07 \end{array}$ | $\begin{array}{c} -13.52 \pm 0.97 \\ -10.47 \pm 0.21 \\ -7.80 \pm 0.62 \\ -10.60 \end{array}$ |
| Y _{III} M ₃ A _{III} | M1/M2 M2/M3 M3/A4 average | $\begin{array}{c} 14.06 \pm 0.72 \\ 16.34 \pm 0.32 \\ 21.84 \pm 1.61 \\ 17.42 \end{array}$ | $\begin{array}{c} -21.47 \pm 0.17 \\ -20.43 \pm 0.26 \\ -28.32 \pm 1.24 \\ -23.41 \end{array}$ | -7.93 ± 0.07 -6.29 ± 0.09 -8.34 ± 0.67 -7.52 |
| Y _{III} M ₃ A _{II} | M1/M2 M2/M3 M3/A4 average | 16.17 15.08 18.76 16.67 | -25.94 -24.64 -23.25 -24.61 | -11.70 -10.91 -5.06 -9.22 |
| importin-α (1bk5) | R6/R7 R7/R8 R8/R9 average | $13.28 \pm 0.68 \\ 19.10 \pm 0.47 \\ 27.43 \pm 0.42 \\ 19.94$ | $\begin{array}{c} -23.96 \pm 0.01 \\ -22.87 \pm 0.48 \\ -27.68 \pm 0.16 \\ -24.84 \end{array}$ | $\begin{array}{c} -13.58 \pm 0.46 \\ -13.83 \pm 0.22 \\ -13.49 \pm 0.23 \\ -13.63 \end{array}$ |
| importin-α: NLS complex (1bk6) | R6/R7 R7/R8 R8/R9 average | $\begin{array}{c} 13.11 \pm 0.19 \\ 18.66 \pm 0.13 \\ 27.76 \pm 0.16 \\ 19.84 \end{array}$ | $\begin{array}{c} -24.64 \pm 0.01 \\ -22.15 \pm 0.09 \\ -27.60 \pm 0.03 \\ -24.80 \end{array}$ | -12.76 ± 0.37 -12.82 ± 0.02 -13.04 ± 0.13 -12.95 |

Table S2. Super-helical parameters of importin- α and designed ArmRPs as defined by Forwood et al.¹

* The PDB IDs are given in brackets.

[#] Given are the repeats that define the indicated super-helical parameters. The designed ArmRP internal repeats (M-type), C-caps (A_{II} or A_{III}-type) and importin- α NLS binding repeats are defined by the following residue ranges: 43-84 (M1), 85-126 (M2), 127-168 (M3), 169-210 (M4), 169-206 (A4), 211-248 (A5), 289-330 (R6), 331-372 (R7), 373-414 (R8), and 415-456 (R9).

[†] Values are given in degrees and were calculated between the indicated repeat pairs. For designed ArmRPs that crystallize with multiple copies in the AU, the values were averaged across individual copies and errors estimates are given. Since $Y_{III}M_3A_{II}$ crystallized with one molecule in the AU no error estimates are given.

| Name | Sequence 5'-3' direction | Description (for=forward, rev=reverse) |
|---------------------|--|--|
| pQE_f_1 | CGGATAACAATTTCACACAG | forward primer for pQE vectors |
| pQE_r_1 | GTTCTGAGGTCATTACTG | revers primer for pQE vectors |
| Y _{II} 1F | CCAGGCATCCGAACTGCCGCAGATGAC CCAGCAGCTGAACTCTG | for assembly Y_{II} module and amplification |
| Y _{II} 2R | CGGTAGCAGACAGCTGTTCCTGCATGTCG TCAGAGTTCAGCTGCTGGG | rev assembly Y_{II} module |
| Y _{II} 3F | GAACAGCTGTCTGCTACCCGTAAATTCTC TCAGATCCTGTCTGATGG | for assembly $Y_{\rm II}$ module |
| Y _{II} 4R | TTCCTGGTACCCTAAGGTCTCAACCATCA GACAGGATCTGAGAG | rev assembly $Y_{\rm II}$ module and amplification |
| $Y_{III}1F$ | CCAGGGATCCGAACTGCCGCAGATGGTTC AGCAGCTGAACTCTC | for assembly $Y_{\mbox{\scriptsize III}}$ module and amplification |
| Y _{III} 2R | GCAGAGCAGACTGCAGTTCCTGCTGGTCC GGAGAGTTCAGCTGCTGAACC | rev assembly Y_{III} module |
| Y _{III} 3F | GAACTGCAGTCTGCTCTGCGTAAACTGTC TCAGATCGCTTCTGGAGG | for assembly Y_{III} module |
| Y _{III} 4R | TTCCTGGTACCCTAAGGTCTCAACCTCCA GAAGCGATCTGAG | rev assembly $Y_{\mbox{\scriptsize III}}$ module and amplification |
| $A_{II}1F$ | CCAGGGATCCTAGGAAGACCTTGGTAACG AACAGAAACAGGC | for assembly A_{II} module and amplification |
| A _{II} 2R | GTTTCTCCAGAGCACCAGCTTCTTTAACA GCCTGTTTCTGTTCGTTACC | rev assembly A _{II} module |
| A _{II} 3F | GCTGGTGCTCTGGAGAAACTGGAACAGCT GCAGTCCCACGAG | for assembly A_{II} module |
| A _{II} 4R | CCTGAGCTTCTTTCTGGATCTTCTCGTTC TCGTGGGACTGCAGC | rev assembly A _{II} module |
| A _{II} 5F | GATCGAGAAAGAAGCTCAGGAAGCTCTGG AGAAGCTGCAGTCCC | for assembly A_{II} module |
| A _{II} 6R | TTCCTGGTACCTCATTAGTGGGACTGCAG CTTCTCCAG | rev assembly A_{II} module and amplification |
| A _{III} 1F | CCAGGGATCCTAGGAAGACCTTGGTAACG AACAGAAACAGGC | for assembly A_{III} module and amplification |
| A _{III} 2R | GAGCCGGTTCAGCACCAGCTTCTTTAACA GCCTGTTTCTGTTCGTTACC | rev assembly A _{III} module |
| A _{III} 3F | GCTGGTGCTGAACCGGCTCTGGAACAGCT GCAGTCCTCCCCG | for assembly A _{III} module |
| A _{III} 4R | CCTGAGCTTCTTTCTGGATCTTCTCGTTC GGGGAGGACTGCAGC | rev assembly A _{III} module |
| A _{III} 5F | GATCCAGAAAGAAGCTCAGGAAGCTCTGG AGAAGATCCAGTCCC | for assembly A _{III} module |
| A _{III} 6R | TTCCTGGTACCTCATTAGTGGGACTGGAT CTTCTCCAG | rev assembly A_{III} module and amplification |

Table S3. Oligonucleotides used for the assembly and cloning of designed armadillo repeat protein genes



Supplementary Figure S1. Size-exclusion chromatography combined with multiangle light scattering (MALS). The absorption at 280 nm and the molecular mass are shown as black and magenta curves, respectively. $Y_{II}M_3A_{II}$ was analysed at 5 mg/ml (a) and 18 mg/ml (b) concentration.



Supplementary Figure S2. Sketch illustrating the definitions of curvature, twist, and lateral bending (adapted from Forwood et al. ¹). R1 and R2 indicate two neighboring repeats of the ArmRP. The three principal axes are: the P1 axis is in the plane of the helices but perpendicular to them (green), the P2 axis is parallel to the helices (cyan) and the P3 axis is perpendicular to axes P1 and P2 (red). Curvature (ϕ) is the angle between two P1 axes of adjacent repeats, twist (τ) is the angle between two P2 axes of adjacent repeats and the lateral bending angle (θ) is the angle between two P3 axes of adjacent repeats.



Supplementary Figure S3. SDS-PAGE after single-step IMAC purification of designed ArmRPs. Constructs with 3 to 6 internal repeats. Lane 1, $Y_{II}M_3A_{II}$ (23 kDa); lane 2, $Y_{II}M_4A_{II}$ (27 kDa); lane 3, $Y_{II}M_5A_{II}$ (31 kDa); lane 4, $Y_{II}M_6A_{II}$ (35 kDa).

References

1. Forwood JK, Lange A, Zachariae U, Marfori M, Preast C, Grubmuller H, Stewart M, Corbett AH, Kobe B (2010) Quantitative structural analysis of importin-beta flexibility: paradigm for solenoid protein structures. Structure **18**:1171-1183.